IBC Meeting Minutes

Chair- Ken Bondioli
Research Safety- Abigail Fish
Thursday, January 15, 2026
1:30 pm via Zoom

Institutions *Louisiana State University Agricultural and Mechanical College (A&M)*
Louisiana State University Ag Center

IBC Members			
	Ken Bondioli	LSU Ag Center	Chair, Animal Expert
	Abigail Fish	LSU A&M	BSO, Administrator, Voting Contact
	Sarah Keeton	LSU A&M	BSO
	Sue Hagius	LSU Ag Center	Animal Expert, Lab Rep
	Michael Hooks	LSU A&M	Member
	Jong Ham	LSU Ag Center	Plant Expert
	Christy White	Pennington Biomedical Research Center	Non-Voting Member
	Jeff Davis	LSU Ag Center	Plant and Insect Expert
	Niranjan Baisakh	LSU Ag Center	Plant Expert
	William Doerrler	LSU A&M	Member
	Ramanuj Lahiri	National Hansen's Disease Program	Member
	Rebecca Christofferson	LSU A&M	Member
	Michelle Dennis	Our Lady of the Lake Hospital	Local Non-Affiliated Member
	Brent Stanfield	LSU A&M	Member
	Ryoichi Teruyama	LSU A&M	Member
	James Bush	Blue Cross Blue Shield of Louisiana	Local Non-Affiliated Member

Members Present:	Ken Bondioli, Abigail Fish, Sarah Keeton, Sue Hagius, William Doerrler, Michael Hooks, Ryoichi Teruyama, Brent Stanfield (arrived at 1:34 pm), Rebecca Christofferson, Ramanuj Lahiri, and Jong Ham.	
Members Absent:	Christy White, Michelle Dennis, Jeff Davis, Niranjan Baisakh, and James Bush.	
Others Present:	Mariano Carossino	Associate Professor, Department of Pathobiological Sciences
	Kristen Healy	Assistant Professor, Department of Entomology
	Xun Tang	Assistant Professor, Department of Chemical Engineering
	Sydney Moyo	Assistant Professor, Department of Biological Sciences
Call to Order:	1:31 pm	
Approval of Minutes from:	January 15, 2026	
	Motion Made by:	Sue Hagius
	Seconded by:	Rebecca Christofferson
	Abstaining:	Jong Ham and Ramanuj Lahiri.

Business and Call for New Business

NIH Noncompliance Report Update: The NIH Office of Science Policy has reviewed the IBC's noncompliance report. The NIH responded that no further action is required at this time. This matter is considered closed unless additional information becomes available.

New IBC Registrations and Amendments for Review

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26003 (Renewal)	Xun Tang	Chemical Engineering	1/28/2026	Design of RNA-Based Circuit for Gene Expression Regulation	Michael Hooks	Ryoichi Teruyama

Project Overview:

This project aims to design, construct, and evaluate synthetic biomolecular circuit networks that regulate target gene expression. The research integrates mathematical modeling with experimental validation to develop predictable gene regulatory systems. Computational models will guide circuit architecture and optimize regulatory interactions prior to laboratory testing. Circuit components will first be characterized in cell-free transcription–translation systems to generate quantitative data for model refinement and feasibility assessment. Finalized gene circuits will then be assembled into plasmid constructs and introduced into laboratory strains of Escherichia coli for functional evaluation. Experimental results will be compared to model predictions to assess circuit performance, stability, and gene expression dynamics.

Risk Assessment and Discussion:

This project presents low biosafety risk consistent with routine recombinant DNA work in non-pathogenic laboratory strains of *Escherichia coli* and the use of acellular transcription–translation systems. The research involves standard molecular cloning, plasmid transformation, and evaluation of synthetic gene circuits.

Potential hazards relate to handling viable *E. coli* cultures and recombinant DNA during routine laboratory procedures, including possible exposure through splashes or aerosol-generating activities. These risks are effectively managed under Biosafety Level 1 (BSL-1) containment through standard microbiological practices, appropriate PPE, surface decontamination, and proper waste disposal.

No human pathogens, viral vectors, toxin genes, or virulence factors are described. With these precautions in place, the project is considered low risk and appropriately contained under standard BSL-1 practices. No environmental or security concerns are anticipated.

NIH Guidelines: Section III-F-8, Appendix C.

Biosafety Level: BSL-1

Training Requirements: All personnel, including the PI, involved in this project must complete BSL-1 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.

IBC Vote: **Approved at BSL-1 pending receipt of modifications**

Motion made by: Michael Hooks

Seconded by: Ryoichi Teruyama

Abstaining: None

Conflicts of Interest: None

Requested Modifications:

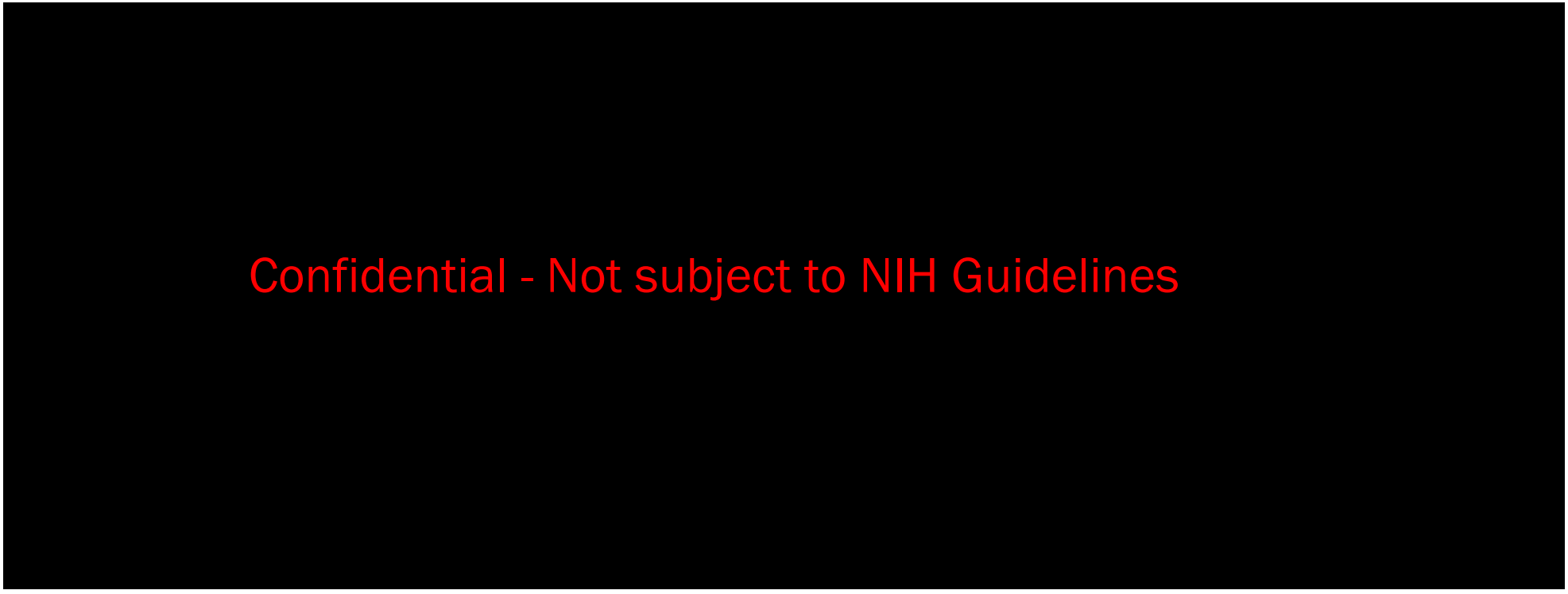
- Section A. Project Information.
  - Locations. Please remove AgMetal building and room information. If you plan to work in that space again in the future, we can add the location as an amendment to this protocol.
- Section B. Project Description.
  - Project Goals. Please describe why regulating gene expression is important to the overall goal of the work.
  - Procedures and Methods. Please indicate which genes you plan to edit and list the microplate reader room number.
- Section C. Risk Evaluation.
  - Biosafety. Please state that you will use the BSC for sterile media preparation and bacterial culture expansion.
  - Biosafety. Please indicate that you will decontaminate liquid waste in a final concentration of 10% bleach for a minimum of 30 minutes before discarding it down the drain. Please add a statement indicating that, if potentially infectious material must be moved between labs, leak-proof primary and secondary containment will be used.

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26004	Kristen Healy	Entomology	2/2/2026	Rearing of Local Calliphoridae Blow Flies in the Lab	Sarah Keeton	Sue Hagius

Project Overview: This project was placed on hold pending receipt of additional information necessary to complete a full review. A project overview is not included at this time due to incomplete documentation.

Risk Assessment and Discussion:	The risk assessment for this project has been deferred pending receipt of additional information necessary to complete a thorough evaluation.
NIH Guidelines:	Not Applicable
Biosafety Level:	To Be Determined
Training Requirements:	To Be Determined
IBC Vote:	**The protocol was placed "on hold" until further information is provided.**
	Motion made by:
	Seconded by:
	Abstaining:
	Conflicts of Interest:

Requested Modifications:



Confidential - Not subject to NIH Guidelines

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26005	Mariano Carossino	Pathobiological Sciences	2/3/2026	Development of Vaccine Strategies Against HPAI H5N1 and Other Viral Pathogens of Poultry	Rebecca Christofferson	Brent Stanfield

Project Overview:

This project aims to develop and evaluate vaccine platforms for highly pathogenic avian influenza (HPAI) H5N1 and infectious bronchitis virus (IBV). The research includes the development of self-amplifying RNA (saRNA) and replication-incompetent vesicular stomatitis virus (rVSV)–vectored vaccines expressing viral antigens associated with these pathogens.

Additional work will involve the development of single-domain antibodies (nanobodies, VHH) targeting immune cell receptors using naïve immune libraries in a yeast display system to support the targeted delivery of vaccine antigens. Vaccine candidates generated in this work will be evaluated for immunogenicity in chickens using in ovo vaccination and drinking water administration methods.

The overall objective is to assess the feasibility and the immune responses generated by these vaccine platforms against avian influenza and infectious bronchitis viruses.

Risk Assessment and Discussion:

This project presents a moderate biosafety risk consistent with the development and evaluation of viral vector and RNA-based vaccine platforms for avian pathogens. The research involves generating replication-incompetent vesicular stomatitis virus (rVSV; Indiana strain) vectors expressing viral antigens, developing self-amplifying RNA (saRNA) vaccine constructs, and using yeast display systems to generate single-domain antibodies for targeted antigen delivery. No live highly pathogenic avian influenza (HPAI) virus is used.

Potential hazards include handling recombinant viral vectors and nucleic acid constructs, and performing laboratory procedures involving avian cells, eggs, and chickens during immunogenicity studies. These risks are effectively managed under Biosafety Level 2 (BSL-2), Animal Biosafety Level 2 (ABSL-2), and BL2-N practices, including appropriate PPE, use of a biological safety cabinet for aerosol-generating procedures, routine decontamination, and proper disposal of biohazardous materials.

With these precautions in place, the project is considered moderate risk but appropriately contained under standard BSL-2, ABSL-2, and BL2-N practices. No environmental or security concerns are anticipated.

NIH Guidelines:

Section III-D-1-a
Section III-D-2-a
Section III-D-3-a
Section III-D-4-b

Biosafety Level: BSL-2, ABSL-2, and BL2-N

Training Requirements: All personnel, including the PI, involved in this project must complete BSL-2 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. In addition, all staff involved in animal procedures or husbandry must complete LSU's approved animal handling and species-specific training as required by the IACUC. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.

IBC Vote: **Approved at BSL-2, ABSL-2, BL2-N pending receipt of modifications**

Motion made by:	Rebecca Christofferson
Seconded by:	Brent Stanfield
Abstaining:	None
Conflicts of Interest:	None

Requested Modifications:

- Section A. Project Information
  - Title. Please spell out HPAI and clarify which other poultry pathogens you plan to use.
  - Locations. Please add room numbers for flow cytometry, live cell sorting, DLAM, media prep, and Genelab as applicable.
- Section B. Project Description.
  - Project Goals. Please rephrase in layman's terms and include a statement about the overarching goal of the research. Please list other poultry pathogens you plan to use. If this list changes, others can be added as amendments at a later time. Please remove mention of llamas. Please describe in ovo vaccination and remove objective 4.
  - Procedures and Methods. Please rephrase in layman's terms and reduce redundancy regarding waste and transport. A sweeping statement at the beginning is sufficient. Please ensure all acronyms are defined at the beginning of this section, specifically the ones related to viral components. Please indicate what work takes place in what room and what work occurs in a BSC; a broad statement at the beginning is fine. Please use state that primary and secondary leakproof containment will be used. Please elaborate on immunofluorescence procedures. Specifically describe how the virus/vaccine strains are inactivated. Please state that VSV Indiana is the wild-type strain. Please state where sequencing is done. If you plan to use Genelab, please add that room number to section A. Please state which generation of lentiviral vector you plan to use and address any additional safety precautions required for working with this viral vector. Please briefly describe cell, viral, bacterial, and yeast culture methods, viral neutralization assays, hemagglutination inhibition assay, and ELISA techniques. Specifically include information on when the pathogen is inactivated. Please add a statement indicating that the VEEV-TC-83 strain has been designated an RG-2 pathogen and that you will not create a replication-competent virus, only RNA containing the genes of interest. Please add details on blood collection and define E18. Please indicate how drinking water is discarded for objective 3. Please add a statement on samples shipped to or received from Kansas state. Please remove objective 4.
- Section C. Risk Evaluation
  - Containment Level. Please uncheck ABSL-1 and BL1-N.
  - Biosafety. Please describe the PPE required for live-cell sorting and animal work. Please also indicate when an N95 is required. Please add the media prep room number to Section A.
  - Biosecurity. Please briefly describe secure transport between labs. Please describe animal carcass use, fixation, and disposal.
- Section F. Recombinant DNA.
  - NIH Guidelines. Please add Section III-D-3-a and Section III-D-4-b.
  - Cells/Organisms. Please list all cell lines used and ensure consistency throughout this section.
  - Vectors. Please add lentiviral vector information to all questions.
  - Experimental Hosts. Please add 1 day old chickens.
- Section K. Animal Pathogens.
  - Pathogens used. Please indicate that VSV Indiana is the wild-type strain.
  - Ultracentrifugation. Please add room number to Section A.
  - Source of Pathogen. Please indicate what pathogens comes from what source.

- Permit. Please indicate which pathogens require a permit under Section B. Procedures.

- Section N. Safety.
  - Administrative Controls. Sharps. Please check all appropriate sharps. Disinfection/Decontamination. Please uncheck autoclave for solid waste.
  - Engineering Controls. BSC. Please update the BSC certification date.
  - Personal Protective Equipment. Please check N95
  - Other Safety Equipment. Please check safety shower and fire extinguisher.

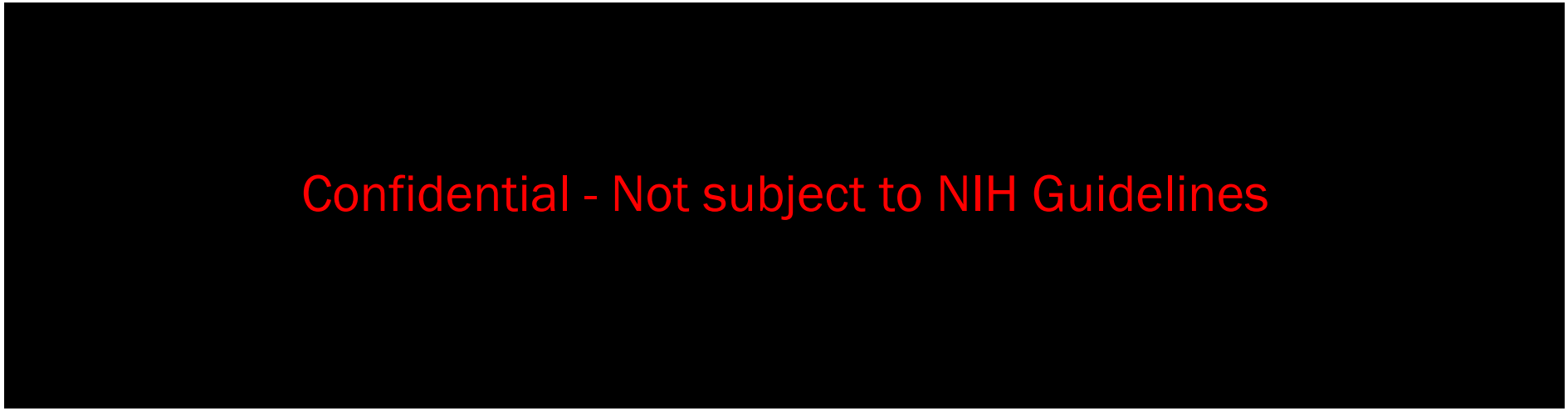
Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26006	Sydney Moyo	Biological Sciences	2/3/2026	LSU Rural Scholars: Mercury in Fishes Consumed by Residents in Rural Communities in Terrebonne and Lafourche Parishes	Sue Hagius	Sarah Keeton

Project Overview:	This project evaluates the effects of climate-driven flooding and industrial activity on fisheries and human health in coastal Louisiana, with a focus on Terrebonne Parish. The study assesses how flooding events influence fish availability and access and examines mercury contamination in commonly consumed fish in relation to oil and gas infrastructure. Ecological sampling and community dietary survey data will be used to estimate mercury exposure within coastal populations. The primary objectives are to: (1) assess the impact of flooding on local fisheries and food security; (2) evaluate mercury contamination in fish; and (3) estimate community mercury burden based on dietary exposure patterns.
Risk Assessment and Discussion:	This project presents moderate biosafety risk consistent with laboratory handling and homogenization of environmental fish tissue specimens. The research involves collection and processing of fish samples for mercury analysis, including tissue dissection and mechanical homogenization, as well as collection of community dietary survey data. Potential hazards relate to handling raw fish tissue that may harbor naturally occurring aquatic pathogens (e.g., *Vibrio* spp. or other environmental microorganisms), particularly during tissue homogenization or other aerosol-generating procedures. These risks are effectively managed under Biosafety Level 2 (BSL-2) containment, including use of biological safety cabinets for aerosol-generating activities, appropriate personal protective equipment (PPE), surface decontamination, and proper biohazard waste disposal in accordance with institutional protocols.

No intentional pathogen culture, recombinant DNA work, or regulated infectious agent manipulation is described. With these precautions in place, the project is considered moderate risk but appropriately contained under standard BSL-2 practices. No environmental or security concerns are anticipated.

NIH Guidelines:	Not Applicable
Biosafety Level:	BSL-2
Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-2 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. In addition, all staff involved in animal procedures or husbandry must complete LSU's approved animal handling and species-specific training as required by the IACUC. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-2 pending receipt of modifications**
Motion made by:	Sue Hagius
Seconded by:	Sarah Keeton
Abstaining:	None
Conflicts of Interest:	None

Requested Modifications:



Confidential - Not subject to NIH Guidelines

Reg. #	PI Name	Affiliation of PI	Date Received	Title of Project	Reviewer 1	Reviewer 2
26007	Levent Dirikolu	Comparative Biomedical Sciences	2/4/2026	Pivotal Field Study to Evaluate the Effectiveness and Safety of SB-001 for the Treatment of Anemia Associated with Chronic Kidney Disease	William Doerrler	Jong Ham

Project Overview:

This project evaluates the safety and biological activity of SB-001, an investigational gene therapy intended for the treatment of non-regenerative anemia associated with chronic kidney disease (CKD) in cats. SB-001 consists of a recombinant adeno-associated virus (AAV) vector containing a DNA transgene encoding feline erythropoietin (fEPO). The vector is administered as a single intramuscular injection with the goal of achieving sustained in vivo expression of fEPO.

The study will assess field safety following intramuscular administration of the recombinant vector and evaluate biological activity through measurement of packed cell volume (PCV) to determine whether treatment results in improvement of anemia. Data generated from this study are intended to support potential regulatory submissions for the development of SB-001 as a veterinary therapeutic.

Risk Assessment and Discussion:	This project presents low biosafety risk consistent with clinical administration of a recombinant adeno-associated virus (AAV) gene therapy product in cats. The study involves intramuscular injection of SB-001 and subsequent clinical monitoring and specimen collection to evaluate safety and biological activity. Potential hazards relate to accidental needlestick or contact with animal tissues or waste that may contain limited amounts of vector following administration. No vector production, propagation, or genetic manipulation occurs at this site; the investigational product is manufactured externally under a separate approved IBC protocol. Risks are managed under BSL-1, ABSL-1, and BL1-N practices, including appropriate PPE, sharps safety, routine decontamination, and proper disposal of animal waste and materials. With these precautions in place, the project is considered low risk and appropriately contained under assigned biosafety levels. No environmental or security concerns are anticipated.
NIH Guidelines:	Section III-D-4-a
Biosafety Level:	BSL-1, ABSL-1, and BL1-N
Training Requirements:	All personnel, including the PI, involved in this project must complete BSL-1 training in accordance with LSU's Environmental Health and Safety (EHS) and Institutional Biosafety Committee (IBC) requirements. In addition, all staff involved in animal procedures or husbandry must complete LSU's approved animal handling and species-specific training as required by the IACUC. All training must be completed before beginning work and refreshed as required by LSU policies and SOPs.
IBC Vote:	**Approved at BSL-1, ABSL-1, and BL1-N pending receipt of modifications.**
	Motion made by: William Doerrler
	Seconded by: Jong Ham
	Abstaining: None
	Conflicts of Interest: None

Requested Modifications:

- Section A. Project Information
  - Buildings. Please add the LADDL building. Room numbers. Please list the small animal examination room numbers.
  - Personnel. Please list EHS-specific training under specific training and describe the experience for all personnel and the PI. Please add Catherine Takawira as a staff member and any other students or technicians who will administer vaccinations.
- Section B. Project Description.
  - Project Goals. Please spell out EPO the first time you use the acronym. Please relocate all information listed under methods at LSU to the procedures and methods.

    - Procedures and Methods. Please indicate a statement indicating that the recombinant vaccine is not manufactured at LSU but is provided by a commercial company. Please indicate which room numbers SB-100 is received, stored, and prepared. Please add a room number for the prep area. Please indicate what disinfectant will be used before and after dosing. Please add a statement on PPE required for students and staff.
- Section C. Risk Evaluation
    - Containment Level. Please check BSL-1.
    - Biosafety. Please add that safety glasses are required PPE, especially during injections. Please also add a statement on the containment of aerosols.
    - Biosecurity. Please indicate that liquid waste is decontaminated in a final concentration of 10% bleach for a minimum of 30 minutes of contact time. Please state that materials will be disposed of in biohazardous waste containers and add a statement on sharps.
- Section N. Safety.
    - Administrative Controls. Please check 10% bleach for liquid waste.
    - PPE. Please check safety glasses.
    - Other safety equipment. Please check fire extinguishers, first-aid kit, phone, and sink/soap.

Upcoming Meetings:	March 12, 2026 @1:30 pm via Zoom
Adjourned:	3:25 pm