

Bottom-up trait-mediated indirect effects decrease pathogen transmission in a tritrophic system

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Abstract. A plant's induction of secondary defenses helps to decrease herbivore damage by changing resource quality. While these chemical or physical defenses may directly decrease herbivory, they can also have indirect consequences. In a tritrophic system consisting of a plant, an insect herbivore, and an insect pathogen, plant based trait-mediated indirect effects (TMIEs) can alter host–pathogen interactions and, thereby, indirectly affect disease transmission. In a series of field experiments, individual soybean plants (*Glycine max*) were sprayed with either a jasmonic acid (JA) solution to trigger induction of plant defenses or a similar control compound. Fall armyworm (*Spodoptera frugiperda*) larvae along with varying amounts of a lethal baculovirus were placed on the plants to measure transmission. Induction of plant defenses decreased viral transmission due to increased population heterogeneity arising from changes in individual susceptibility. The change in susceptibility via TMIEs was driven by a decrease in feeding rates and an increase viral dose needed to infect larvae. While the induction against herbivore attack may decrease herbivory, it can also decrease the efficacy of the herbivore's pathogen potentially to the plant's detriment. While TMIEs have been well-recognized for being driven by top-down forces, bottom-up interactions can dictate community dynamics and, here, epizootic severity.

Key words: *Baculovirus*; *Bayesian analysis*; *epizootics*; *fall armyworm*; *induced plant defenses*; *Spodoptera frugiperda*; *variability in transmission rate*.

INTRODUCTION

In tritrophic systems, trait-mediated indirect effects (TMIEs) occur when changes in a species trait directly alters the interactions between two trophic levels and indirectly affects the third (Werner and Peacor 2003). From a top-down perspective, non-consumptive effects (NCEs) between predator and prey can initiate this cascade. NCEs happen when the mere presence of a predator changes prey behavior, thereby affecting the prey's development, growth, or survival (Peckarsky et al. 2008). NCEs can, in fact, be even larger than those dictated by changes due to direct predator consumption of the prey (Bolker et al. 2003, Preisser et al. 2005). In turn, changes in the prey's behavior can have cascading consequences on the resources that the prey consume resulting in a TMIE between the predator and the basal resource in the system. Most work on TMIEs focuses on how predator presence changes prey dynamics via this top-down perspective (Beckerman et al. 1997, Peckarsky et al. 2008, Schmitz et al. 2016). From a disease ecology perspective, this can lead to changes in infection risk (Buck and Ripple 2017). Yet, increasing evidence shows that bottom-up processes, due to changes in resource type and resource quality via TMIEs, may be equally important for determining transmission dynamics (Hall et al. 2009, Elderd et al. 2013, Tao et al. 2015).

Plants, as basal resources in a tritrophic system, may change the chemical composition of their leaves via induced defenses to decrease herbivore consumption (Karban and

Baldwin 1997). The advantage of employing an induced defense is that induction reduces the cost of the defense as compared to constitutive defenses and that increasing variability in the resource can lead to a decline in herbivore performance (Karban et al. 1997). For specialists, chemical defenses, either constitutive or induced, can be sequestered as an anti-predator defense (Bowers 1980, Agrawal et al. 2012) or detoxified (Ratzka et al. 2002), which serves little benefit for the plant. While for generalists, changes in resource quality due to induction may decrease herbivore performance, induced plant defenses can have important indirect effects on upper trophic levels and may change the interaction between a predator and its prey (Ohgushi 2012, Kersch-Becker and Thaler 2015) as well as between a host and its pathogen (Hall et al. 2009, Elderd et al. 2013). Thus, changes in plant resource quality can have cascading consequences for other actors in the system via TMIEs (Werner and Peacor 2003, Ohgushi et al. 2012, Stephens et al. 2017). For instance, insect host–pathogen interactions may be dictated by the resource quality of the plant on which the insect feeds such that resource quality determines whether the insect becomes infected or not (Hunter and Schultz 1993, Cory and Hoover 2006, Elderd et al. 2013).

To determine how changes in resource quality affect host–pathogen interactions, I used a tritrophic system consisting of a lethal insect virus, an herbivore, and a plant variety that varies in its production of secondary chemical defenses (Shikano et al. 2017). Using both field and laboratory experiments, I show that resource quality changes disease transmission by increasing the variability in host susceptibility. This results in a decrease in epizootic severity at higher virus levels when plant defenses have been induced. Thus, while plant defenses may decrease herbivory, they can also decrease pathogen efficacy potentially to the plant's detriment.

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METHODS

Study system

A species-specific baculovirus, *Spodoptera frugiperda* multicapsid nucleopolyhedrovirus (SfMNPV) represents the upper trophic level in this insect pathogen, herbivore, and plant system. Baculoviruses are ubiquitous in nature and infect a wide-range of insect species (Miller 1997). Epizootics begin when foliage contaminated with baculovirus occlusion bodies (OBs) are consumed by a susceptible larva (Cory and Myers 2003). The OBs contain multiple virions surrounded by a protein coat, which dissolves in the host midgut. If enough OBs are consumed, a fatal infection occurs. The virus then replicates within the host until the baculovirus triggers the host's liquefaction. While uninfected individuals continue to molt to larger instars, infected individuals do not. Horizontal transmission occurs when OBs are released and contaminate the foliage on which susceptible larvae feed (ElderD 2013). Over time, the virus particles degrade due to UV light exposure.

The polyphagous fall armyworm, which serves as the host for SfMNPV, is a multivoltine migratory species with non-overlapping generations that feeds on a number of different crop plants (Sparks 1979, Richter et al. 1987). Adults annually migrate to the Southern United States in April and May from Florida and Texas, where the species overwinters, and continue northward in subsequent generations. Adult females lay eggs in clusters of up to a few hundred (Sparks 1979). After the eggs hatch, there are six larval instars before pupation (Pitre and Hogg 1983). Outbreaks of the fall armyworm, which have been recorded as early as 1845 (Hinds and Dew 1915), can be quite large and wide-spread and can result in as many as 59 pupae/m² (Pair et al. 1991). Additionally, the armyworm/spodoptera complex does substantial damage to soybean and other crops in the Southeast (Musser et al. 2018). The fall armyworm is a particular problem in the soybean fields of Brazil (Peruca et al. 2018). Recently, *S. frugiperda* has been introduced into Africa and is causing widespread damage (Stokstad 2017). For the fall armyworm, SfMNPV infection rates are quite high, reaching 50–60% in infested areas (Fuxa 1982). Thus, SfMNPV represents an important source of mortality (Richter et al. 1987).

Soybean isolines, the basal resource in this tritrophic system, vary in the amounts of constitutive and induced defenses that they produce (Bi and Felton 1995, Underwood et al. 2000). Feeding by chewing insects on soybean leaves triggers a large number of defense-related changes locally and systemically (Underwood et al. 2002, Shikano et al. 2017). For the fall armyworm, which readily feeds on soybeans (Sparks 1979, Richter et al. 1987), the production of plant defense compounds may have varying effects (Pitre and Hogg 1983, Paulillo et al. 2000). Given the wide-diet breadth of *S. frugiperda*, the herbivore could be considered a generalist and its response to *G. max* defenses may be indicative of a wide-range of species that feed upon the plant. Additionally, larval mass gain on the soybean isolate used in these experiments, Stonewall, declines when the plants have been induced as does leaf area eaten (Shikano et al. 2017). These differences between feeding on induced

and non-induced plants were not driven by changes in total phenolics or peroxidase activity. Most likely, the differences in performance and consumption were driven by the chemical composition of the induced leaf tissue (Shikano et al. 2017). Overall, these induced changes affect insects directly by altering feeding behavior, damaging midgut tissues, and interfering with digestive processes (Shikano 2017). These changes may also affect insects indirectly by interacting with baculoviruses in insect midguts (Hoover et al. 1998, ElderD et al. 2013).

Population-level dynamics

The baculovirus transmission cycle governing a single outbreak or epizootic can be described by a series of differential equations (Dwyer et al. 1997, Reilly and ElderD 2014) that track the number of susceptible individuals, the infected individuals, and the pathogen in the system. The equation for the change in susceptible individuals over time t takes the form

$$\frac{dS}{dt} = -\bar{\beta} \left[\frac{S(t)}{S(0)} \right]^{C^2} SV \quad (1)$$

where $\bar{\beta}$ is the mean transmission rate and C is the coefficient of variation given the mean transmission rate. S and V are the number of susceptibles and the amount of virus in the system, respectively. During an epizootic, the transmission rate $\bar{\beta}$ is scaled by the fraction of susceptibles in the population at time t and the number of susceptibles at the beginning of the epizootic, $[S(t)/S(0)]$. The degree of scaling is determined by C^2 . This transmission scaling factor, $[S(t)/S(0)]^{C^2}$, starts at one and declines over time resulting in lower transmission as the epizootic progresses (ElderD and Reilly 2014). The above equation assumes that disease transmission is governed by a distribution with a mean and some variation about the mean. At the beginning of the epizootic, transmission is relatively high as the highly susceptible individuals become infected first and is governed solely by the mean transmission rate $\bar{\beta}$. As the epizootic begins to wane, the only uninfected individuals left have a low susceptibility to the disease and overall transmission drops rapidly or slowly depending upon the degree of heterogeneity in the population (See Dwyer et al. 1997: Fig. 1A). As the coefficient of variation decreases to zero, such that all individuals are equally susceptible, the above equation becomes $dS/dt = \beta SV$ (Dwyer et al. 1997). For Eq. 1, either the mean transmission rate and/or the coefficient of variation can differ given the induction status of the plant. That is, the resource quality of the plant as determined by a trait-mediated induced defense can indirectly affect transmission dynamics.

Eq. 1 can be integrated from time 0 to time T , where T corresponds to either a point of time in the epizootic or the end of an experiment. The integrated equation is

$$\frac{S(T)}{S(0)} = (1 + C^2 \bar{\beta} V(0) T)^{-1/C^2} \quad (2)$$

where $S(T)/S(0)$ is the fraction of larvae that survive at the end of the experiment and $V(0)$ is the virus-killed cadaver

density at the beginning of the experiment. The above equation can be easily fit to field data to estimate both the mean transmission rate $\bar{\beta}$ and the associated coefficient of variation C . If instead, there is no heterogeneity in the system and all individuals are equally susceptible, the corresponding differential equation integrates to $S(T)/S(0) = \exp(-\beta V(0)T)$ and only the transmission rate β needs to be estimated.

To understand how up regulation of plant secondary metabolites affects disease transmission, a series of experiments were conducted that manipulated plant induction status and virus-killed cadaver density. Induction was initiated by spraying either a solution containing 1 mmol/L of jasmonic acid (JA) dissolved in ethanol or distilled water on a group of similar sized soybean plants from the inducible Stonewall isoline (Underwood et al. 2000, 2002) every other day for 1 week. The period was chosen to ensure the defenses were induced over the course of the entire field experiment given that levels of defenses in soybeans can decline after 72 h when exposed to herbivores (Underwood et al. 2000) and was comparable to other long-term studies on soybeans (Accamando and Cronin 2012). Another group of plants were sprayed with a control solution that did not contain JA. Using herbivores rather than JA to induce the plants would present an additional challenge as it is often difficult to control for the amount of leaf material consumed across replicates (Baldwin 1996, Cipollini et al. 2003). Soybean plants were germinated and grown in a walk-in chamber at 28°C until being translocated to the field (LSU's Burden Center, Baton Rouge, Louisiana, USA). Virus density was manipulated by placing a varying number of infected first instars (0, 10, 20, or 40) on the plant. An infected first instar will produce 1.1×10^5 baculovirus occlusion bodies (95% Credible Interval [CI], $[0.8 \times 10^5, 1.3 \times 10^5]$) when the instar succumbs to the virus. By placing first instars on the plant rather than spraying virus, the virus would be distributed across the leaf tissue closer to what occurs naturally. To infect the first instars, 9 μ L of 10^6 OBs per 3 μ L solution were spread across two-ounce cups filled with an artificial diet (Southland Productions, Lake Village, Arkansas, USA). Recently hatched first instar larvae (Benzon Research, Carlisle, Pennsylvania, USA) were placed on the diet cup and allowed to feed. Infected larvae do not molt to the next instar and, thus, are easily identified. The infected larvae were then placed on soybean plants covered in a mesh bag to ensure that the larvae did not escape. The mesh bag also prevents virus degradation due to UV light exposure (Elder and Reilly 2014). After four days, during which the first instars had died, 20 healthy fourth-instar larvae were placed on each plant and allowed to feed for an additional four days. The fourth instars were then removed and placed on individual diet cups where they were reared until they had either pupated or died. Recovery varied between replicates due to healthy fourth instars being cannibalized when the mesh bag was opened for recovery. Cannibalism occurred during recovery when the larvae dropped from the leaves and fell to the bottom of the experimental bag covering the plant. When confined to a small space, the larvae would begin consuming nearby conspecifics. Death due to viral infection was confirmed by either liquefaction of the host or under a light microscope where OBs are visible (Cory and Myers 2003).

A total of 20 replicates were used across all control and infected instar treatments. In the plots where no virus was added, no infected individuals were recovered. Thus, there was no need to correct the data for background infection (Morgan 1992) due to environmental contamination. Additionally, given the lack of infection in the zero virus treatments and that having no virus in the system would result in zero transmission, the estimates for the linear and nonlinear models were placed through the origin. The data collected were then fit to the nonlinear heterogenous equation (Eq. 2) or its linear counterpart. The models (Table 1) tested whether the nonlinear or linear transmission model best fit the data depending on plant induction treatment and, thus, whether or not TMIEs via induction of secondary metabolites affected transmission dynamics.

Individual-level dynamics

While the above focuses on the epizootic from a population perspective, it is also useful to examine how TMIEs affect individual probability of infection. In baculovirus-driven systems, an individual larva has to both consume a lethal viral dose and become infected (Dwyer et al. 2005, Elder et al. 2008) in order for horizontal transmission to occur. This process encompasses the two main components of transmission, which include contact rate and infection probability given contact with the virus (Hawley et al. 2011, McCallum et al. 2017). The probability of consumption and infection can be written as $P(I, C) = P(I|C)P(C)$ where $P(I, C)$ represents the joint probability of infection I and virus consumption C . $P(C)$ is the probability of consumption and $P(I|C)$ is the conditional probability of infection

TABLE 1. WAIC and DIC values for the six models considered for the field transmission data, the four models for the dose-response experiments, and the two models for the preference experiment.

Model	WAIC values	DIC values
Field transmission experiment		
1. Linear, no difference	48.98	49.09
2. Nonlinear, no difference	48.96	49.11
3. Both linear	50.71	50.71
4. Both nonlinear	46.74	47.33
5. JA nonlinear, control linear	46.76	47.34
6. JA linear, control nonlinear	50.69	50.70
Dose response experiment		
1. No difference	134.40	140.51
2. Intercept only differs	129.43	129.33
3. Slope only differs	129.59	131.42
4. Both intercept and slope differ	132.16	133.18
Preference experiment		
1. No difference	765.72	763.31
2. Treatment difference	722.38	718.84

Notes: For the field transmission data, the two best-fit models are in bold. Nonlinear and linear refer to the solved heterogenous equation (Eq. 2) and its linear counterpart that assumes that individuals do not differ in susceptibility. The first two models for the field transmission data considered assume no difference in JA-induced and control plants. For the dose-response experiments, the four models considered include a null model of no differences and three models where either the intercept and/or the slope differs between JA-induced and control plants. The best-fit model is highlighted for the dose-response and preference experiments.

given consumption. $P(I|C)$ and $P(C)$ essentially break down the transmission process into two separate actions that incorporate individual behavior and physiology.

To test whether the probability of infection given consumption, $P(I|C)$, changes due to plant induction status, a dose-response experiment was performed. In the experiment, a known amount of virus, either 0, 300, 3,000, or 30,000 OBs, was placed on a 9.5 mm diameter leaf disk from either induced or non-induced plants. Thirty recently molted fourth-instar larvae that had been starved for 24 h were presented with a leaf disk containing one of the experimental viral doses. Only larvae that consumed the entire leaf disk were included to ensure that the larvae received a full dose of the virus and the associated secondary plant metabolites. The number of larvae (mean \pm SD) that failed to consume the entire disk did not vary by induction treatment (control discs 5.0 ± 0.82 larvae, JA-treated discs 5.8 ± 0.50 larvae). A Bayesian Poisson t test using methods outlined in *Statistical analysis* showed that there was an increase in the number of individuals that failed to consume the JA-treated leaf disc but that the estimate of this increase was centered around zero given the 95% CIs (0.13, [-0.461, 0.763]). After consuming the leaf disk and the virus, the larvae were placed on individual diet cups and reared to pupation or death. Death resulting from a viral infection was confirmed using the same techniques as the field experiment. None of the controls became infected during the course of the dose-response experiment. Thus, there was no need to correct the dose-response data for background infection (Morgan 1992) due to laboratory contamination. The data were then analyzed using a logit regression (Collett 2003) where either the slope and/or intercept term differed due to induction treatment. In estimating the slope and intercept associated with the logit function only the non-zero dose data were used as is common practice since the logit function is undefined at zero (Collett 2003). The dose-response experiment directly tested whether TMIEs due to resource quality affected the likelihood of transmission given contact between the host and virus.

To quantify the probability of consumption, $P(C)$, a preference test was performed. Individual fourth-instar larvae were placed in a petri dish with two pre-measured 9.5 mm diameter leaf disks, one of which was from an induced plant and the other from a control plant. The larvae were allowed to feed for up to four hours and the disks were remeasured and amount eaten calculated. All measurements were conducted using ImageJ (Schneider et al. 2012). The analysis of the model associated with fall armyworm preference included a random effect for each individual given the experimental setup. While the above experiment does not directly measure consumption, it does serve as a proxy for how secondary plant metabolites affect feeding behavior of the host.

Statistical analysis

All analyses were conducted using a Bayesian framework with vague priors. The individual-level data from the preference test and the dose-response experiment were analyzed using JAGS (Plummer et al. 2003) and the R2JAGS package (Yu-Sung and Masanao 2015). JAGS (Plummer et al. 2003) was also used to test for differences in cannibalism rates in

the field between induced and control treatments when recovering the fourth instars since insect cannibalism rates are known to vary depending upon the level of induced defenses produced by the plant (Orrock et al. 2017). A Poisson t test with vague priors was used to quantify the difference between the control and induced treatments. For the population-level field transmission data, a Bayesian Markov chain Monte Carlo (MCMC) was constructed in R (R Core Team 2015). For each of the analyses, three chains were run from different starting points. The first 10,000 draws were removed to account for the transient dynamics at the start of the chain. The remaining 100,000 MCMC draws were retained to ensure precise estimates of the associated parameters (Link and Eaton 2012). After visually inspecting the chains, multiple tests for convergence were used including the Gelman-Rubin and the Hiedelberg-Welch test (Gelman et al. 2014). The chains were then combined to construct a posterior distribution. Afterward, a posterior predictive check was performed and Bayesian P values were calculated to determine whether the model being considered fit the data collected (Gelman et al. 1996). Bayesian P values near 0.50 indicate that the model does a reasonable job of fitting the data considered (Kéry 2010). All of the models considered passed each of the individual tests. The associated Watanabe Akaike Information Criteria (WAIC) and Deviance Information Criterion (DIC) scores were calculated (Hobbs and Hooten 2015). Model comparisons were done by comparing both the WAIC and DIC scores across models. The model with the lowest score represented the best fit model given the data.

RESULTS

The best fit models used to measure population-level dynamics in the field showed that the transmission process differed between induction treatments (Table 1). For the best-fit model, transmission was nonlinear for both induced and non-induced plants due to differences in both mean transmission $\bar{\beta}$ (JA-induced, 0.144 [0.056, 0.374]; Control, 0.004 [0.003, 0.006]; median [95% CI]) and the heterogeneity parameter C (JA-induced, 4.719 [3.661, 6.131]; Control, 0.007 [0.005, 0.012]). Note that the estimates of the coefficient of variation for the control treatments were extremely low and very close to zero. As C approaches zero, the nonlinear fit becomes linear and, in this instance, the nonlinear model for the control treatment was essentially linear. The second highest ranking model, which had very similar scores as compared to the best-fit model, also showed that transmission dynamics differed due to induction treatments (Fig. 1). These differences arose from whether the linear or nonlinear model was used to fit the data. JA-induced plants showed decidedly nonlinear transmission. In contrast, host-pathogen transmission dynamics on control plants were decidedly linear. At lower viral loads on the JA-induced plants, transmission increased more rapidly due to an increase in the transmission rate as is to be expected given the heterogeneity model (Dwyer et al. 1997). However, at higher viral loads, transmission declined due to the increase in population-level heterogeneity arising indirectly from the plant's induction of secondary defenses. When comparing the ratio of percent survival on induced vs. non-induced plants, under low viral/cadaver loads survival is less on the

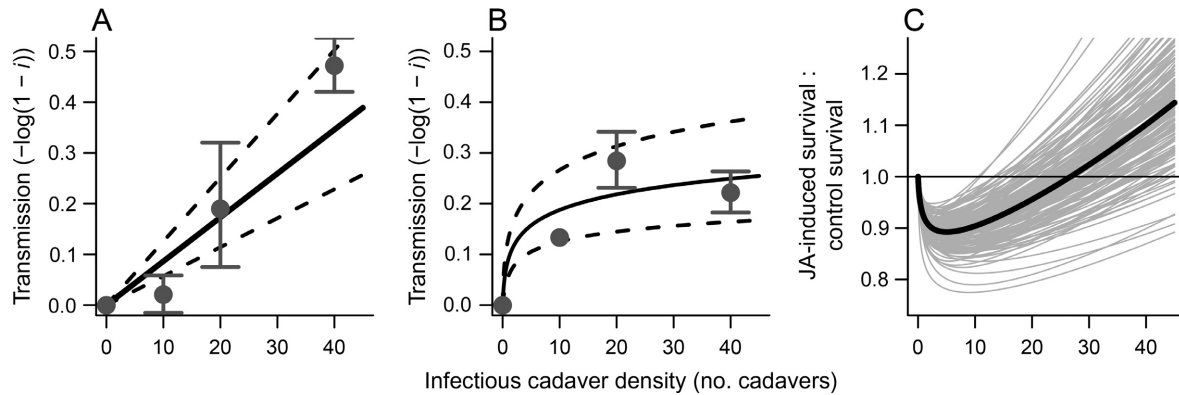


FIG. 1. The effects of increasing infectious cadaver density on population-level transmission on (A) control plants and (B) plants sprayed with jasmonic acid (JA) to induce plant defenses. Transmission, as measured on the y -axis, is the negative log of $1 - i$, where i represents the portion of the individuals infected with the virus. The points represent the means along with their standard deviation. The solid line is the best fit model with 95% credible intervals represented by the dashed lines. Note no standard deviation bars are presented for the 10 cadaver JA treatment since only a single plot was used in the analysis. (C) measures the ratio of percent survival between JA-induced and control field experimental units. The thick dark line is the median ratio and the grey lines are 250 randomly chosen parameter sets to provide an idea of variation about the median. The thin black line at one represents the cross-over point where below the line JA-induced treatments resulted in lower survival. Above the line at higher cadaver levels, JA-induced treatments resulted in higher survival when exposed to the virus as compared to virus on control plants.

induced plant. However, under higher viral loads, induction increases survival (Fig. 1C). Overall, host–pathogen transmission and host survival were governed by changes in resource quality due to bottom-up TMIEs.

For the fourth instars recovered at the end of the field experiment, there were no difference in cannibalism rates. The median difference between cannibalism on induced plants as compared to control plants was positive, albeit rather small, and the 95% CIs overlapped zero (0.07 [−0.428, 0.587]; Appendix S1: Fig. S1).

In terms of individual-level dynamics, there were differences in both the dose-response and preference tests due to the induction treatment (Fig. 2). For the dose response, the best-fit model included differences in the intercept term. Although the second ranked model, which included differences in the slope term, had a relatively similar score (Table 1). The difference in the intercept shifted the curve for JA-induced plants to the right (Fig. 2A,B) such that the lethal dose at which 50% of the population would be infected was much higher for larvae fed virus on JA-induced plants than control plants (Fig. 2C). See Appendix S1: Fig. S2 for the associated logit plot of the relationship. The general result did not change when taking the second ranked model into account (Appendix S1: Fig. S3). The logit model can also be used to calculate a coefficient of variation associated with the dose-response experiment, which can be used as a measure of variability (Elder et al. 2013). Given the logit model's estimates of the intercepts, there was considerable overlap between treatments in the coefficient of variation for the dose-response experiments (JA-induced, 0.196 [0.147, 0.274]; Control, 0.226 [0.168, 0.323]). In terms of larval preference, fall armyworm larvae clearly preferred to feed on the control leaf disk as compared to the JA-induced leaf disk (Fig. 2D, Table 1). Thus, induction of plant secondary metabolites decreased both the $P(I|C)$ and the $P(C)$ for individual larvae.

DISCUSSION

Trait-mediated indirect effects driven by changes in the basal resource clearly affected transmission dynamics. However, the change in dynamics was not entirely due to changes in the mean transmission rate but arose from changes in the variability about the transmission rate. This led to a decrease in infection risk as viral density increased (Fig. 1). From a direct benefit/cost perspective, there are clear direct benefits to the induced defense as fall armyworm feeding declines when defenses are induced (Shikano et al. 2017). There are also clear costs to induction in terms of biomass and seed production (Accamando and Cronin 2012). The indirect costs stem from increasing herbivore survival in the presence of a lethal virus when viral and herbivore densities are high. At high herbivore densities, increased survival due to a decrease in transmission (Fig. 1) will increase defoliation, which will further exacerbate biomass loss. Given the estimates for the transmission rate and its associated CV along with the fact the 50–60% of the population becomes infected during an epizootic (Fuxa 1982), the estimate of virus load or cadaver number can be readily calculated. The median number of first-instar cadavers needed to result in these infection levels is between 90 and 120, which is well above the point where induction of defenses results in lower infection rates as compared to control plants (Fig. 1C). While the induction of chemical defenses may have a direct positive effect on the plant by decreasing herbivory (Karban and Baldwin 1997), the TMIE by decreasing transmission negatively affects the plant and, thus, the induction of the defense in a tritrophic system may do more harm than good from the plant's perspective.

While TMIEs are increasingly recognized as important for driving species dynamics (Bolker et al. 2003, Bolnick and Preisser 2005), the focus of the research often centers on shifts in the mean population response (Flick et al. 2016) and how responses are driven by top-down forces (Beckerman

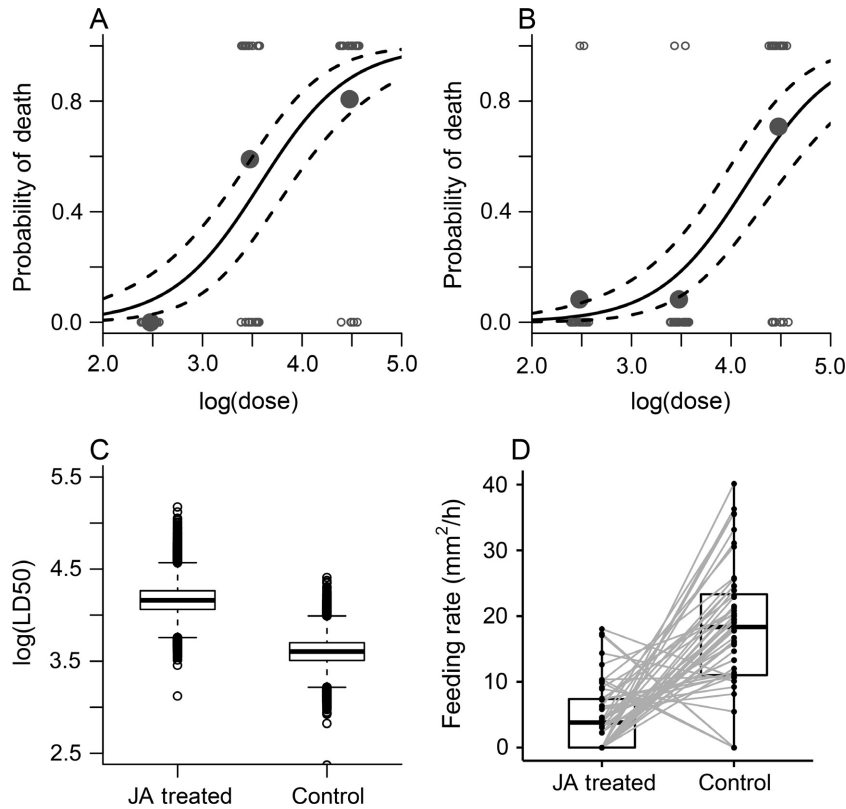


FIG. 2. The effects of plant induction on individual-level dose-response and preference experiments. For the dose-response experiment, the effects of increasing baculovirus dose on the probability of death given the feeding of baculovirus on (A) control leaf tissue and (B) jasmonic acid treated leaf tissue along with the corresponding (C) box plot of the lethal dose at which 50% of the individuals would be expected to die (LD50). For panels A and B, the large solid points represent the mean response for each dose and the small open points are the individual data. These data are jittered for ease of presentation. (D) Box plots of the feeding rate of larvae on JA-treated and control leaf disks during the preference experiment. The small points connected with gray lines represent the paired measurements for individual larvae. In the box-and-whisker plots, the mid-lines are median values, the box limits show the first and third quartiles, the whiskers extend beyond the box limits by 1.5 times the interquartile range, and the points beyond the box limits are outliers.

et al. 1997, Peckarsky et al. 2008). Yet, these effects may not solely impact the mean response but also variability about the mean. For disease transmission, changes in the variability may be driven by either variability in the contact rate or infection risk after contact. While infection risk decreases after induction, there was no measurable change in the CV given the best-fit dose-response model (Table 1, Fig. 2). On the other hand, feeding behavior, which influences contact rate, may be driving the pattern. However, feeding rates were not directly measured in the preference test conducted. Yet others have shown that the feeding rate of fall armyworm declines when soybean defenses in the same isolate used in these experiments are induced (Shikano et al. 2017). Regardless of the exact mechanism, the increase in variability can have important consequences for both the short-term epizootic and, potentially, the long-term population dynamics of the host (Elder et al. 2013).

For the fall armyworm–soybean system, the field transmission results contrast transmission dynamics observed in a baculovirus system where the insect host is the gypsy moth (*Lymantria dispar*) and the virus is consumed on JA-induced or non-induced red oak (*Quercus rubra*). For the gypsy moth, baculovirus transmission on non-induced branches resulted in a nonlinear model best fitting the data and the JA-induced branches resulted in linear transmission (Elder et al. 2013).

The difference between the two systems likely arises because the plants involved produce very different defenses when confronted with an herbivore. When induced, oaks increase the production of hydrolyzable tannins (Hunter and Schultz 1993). Soybeans, in contrast, produce protease inhibitors and peroxidases (Underwood et al. 2002, Shikano et al. 2017). For the gypsy moth, increases in tannins increased leaf consumption (Elder et al. 2013) as they readily consume leaves that have increased tannin concentrations (Dwyer et al. 2005). An increase in tannins also increased the amount of virus that the larval host needed to consume in order to become infected (Elder et al. 2013). The increase in consumption increased the probability that a host contacted the pathogen but induction decreased the probability of infection given consumption. For the fall armyworm, induction of soybean defenses decreases larval consumption and interferes with larval mass gain (Shikano et al. 2017). It also increased the amount of virus needed to infect a larva (Fig. 2C). The similar shifts in the dose-response curves may stem from how the different defenses impact infection probability within the gut of the host. Further research needs to be done to pinpoint the exact mechanism. Given the similar shift in dose-response curves in the two systems, the changes in dynamics most likely arise from differences in consumption. The increase in consumption that was observed in the

gypsy moth may have ensured that all individuals contacted the virus, thereby reducing or eliminating the inter-individual differences or heterogeneity that result in nonlinear transmission. The reduction in consumption that was observed in the fall armyworm (Shikano et al. 2017), which corresponds to the preference data (Fig. 2D), may have increased differences among individuals, thereby strengthening transmission nonlinearities. Thus, the indirect effect mediated by the plant's induction status depends upon the defense being induced and the herbivore's response.

With regard to how secondary metabolites change resource quality and, subsequently, affect epizootic severity, a direct measurement of secondary chemical metabolites was not conducted for this study. However, it is well known that induction of secondary metabolites in soybeans affects the production of foliar phenolics and their composition within the leaf tissue (Shikano et al. 2017). Induction of these defenses can be costly for the plant as it reduces biomass, seed set, and germination (Accamando and Cronin 2012). Indirectly, these secondary metabolites may decrease baculovirus efficacy by interfering with the dissolution of the outer protein coat of the baculovirus, which is needed in order for the infectious viral particles to be released in the insect's midgut (Felton and Duffey 1990). The compounds may also increase the sloughing rate of the midgut along with any infected midgut cells, which decreases the likelihood of a systemic infection occurring (Hoover et al. 2000). In general, the induction of secondary metabolites within the plant's leaf tissue can result in a decrease in infection risk both in the lab and the field. While the focus of these experiments was on the direct and indirect effects of induced plant defenses on the host–pathogen interactions, there are other avenues by which induced defenses may affect disease transmission. The induction of plant defenses can increase cannibalism rates (Orrock et al. 2017), which, according to theory, may either increase (Sadeh and Rosenheim 2016) or decrease transmission (Van Allen et al. 2017). Since baculoviruses stop infected larvae from molting (Elder 2013), the pathogen affects the population's size structure, where healthy larvae are much larger than infected larvae (see Van Allen et al. 2017: Fig. 4). This change in size structure occurs in a number of different parasite systems (Harrison and Hoover 2012). For the fall armyworm, cannibalism rates increase as the difference in size increases (B. G. Van Allen and B. D. Elder, *unpublished data*). Since only 10% of size-based cannibalistic encounters between healthy fourth-instar larvae and infected first-instar larvae result in infection, cannibalism in a size-structured population can decrease transmission (Van Allen et al. 2017). On the other hand, cannibalism may allow for the parasite to infect a new host that contains a greater amount of resources than the current host. In this instance, cannibalism benefits the parasite (Sadeh and Rosenheim 2016). Given that induction of plant defenses can change cannibalism rates (Orrock et al. 2017) and can affect disease transmission (Sadeh and Rosenheim 2016, Van Allen et al. 2017), these two processes may well interact to influence transmission in systems where they both arise. In the field experiments reported here, there was no difference in the number of individuals cannibalized between induced and control plants due to cannibalism (Appendix S1: Fig. S1). However, cannibalism was not size structured as all

individuals were the same instar. How cannibalism affects transmission dynamics by either increasing or decreasing transmission represents a logical avenue to explore experimentally (Orrock et al. 2017, Van Allen et al. 2017).

On a larger spatial scale, since induction changes resource quality, it can affect herbivore movement (Underwood et al. 2005) and may even result in hypermobility of the infected host (Vasconcelos et al. 1996, Goulson 1997). This, in turn, may affect the spatial spread of the disease (Dwyer 1992). The potential interaction between induced plant defenses, herbivore movement, and the herbivore's infection status also represents a ripe avenue for further exploration.

Due to changes in the secondary metabolites, induced plant chemical defenses indirectly decreased baculovirus efficacy. Therefore, in a tritrophic system, the production of these defenses could indirectly have a deleterious effect on the plant. To determine the magnitude of this effect, future research needs to quantify the demographic costs associated with the TMIE from the plant's perspective. While the research examined the differences in outcome during a single epizootic, the long-term dynamics and the potential consequences for host population cycles are less apparent but have been shown in other systems (Elder et al. 2013). These questions not only arise when thinking about temporal issues but also when considering how pathogen–host–resource dynamics are affected at a larger spatial scale as related to changes in the phytochemical landscape (Elder et al. 2013, Hunter 2016). While the focus on TMIEs often comes from a top-down perspective, the importance of bottom-up interactions dictated by resource quality is clearly evident and can have wide ranging consequences.

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SUPPORTING INFORMATION

Additional supporting information may be found in the online version of this article at <http://onlinelibrary.wiley.com/doi/10.1002/ecy.2551/supinfo>

DATA AVAILABILITY

Data are available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.p9d27nh>

Bret D. Elder. Bottom-up trait-mediated indirect effects decrease pathogen transmission in a tritrophic system. *Ecology*. 2018.

Appendix S1

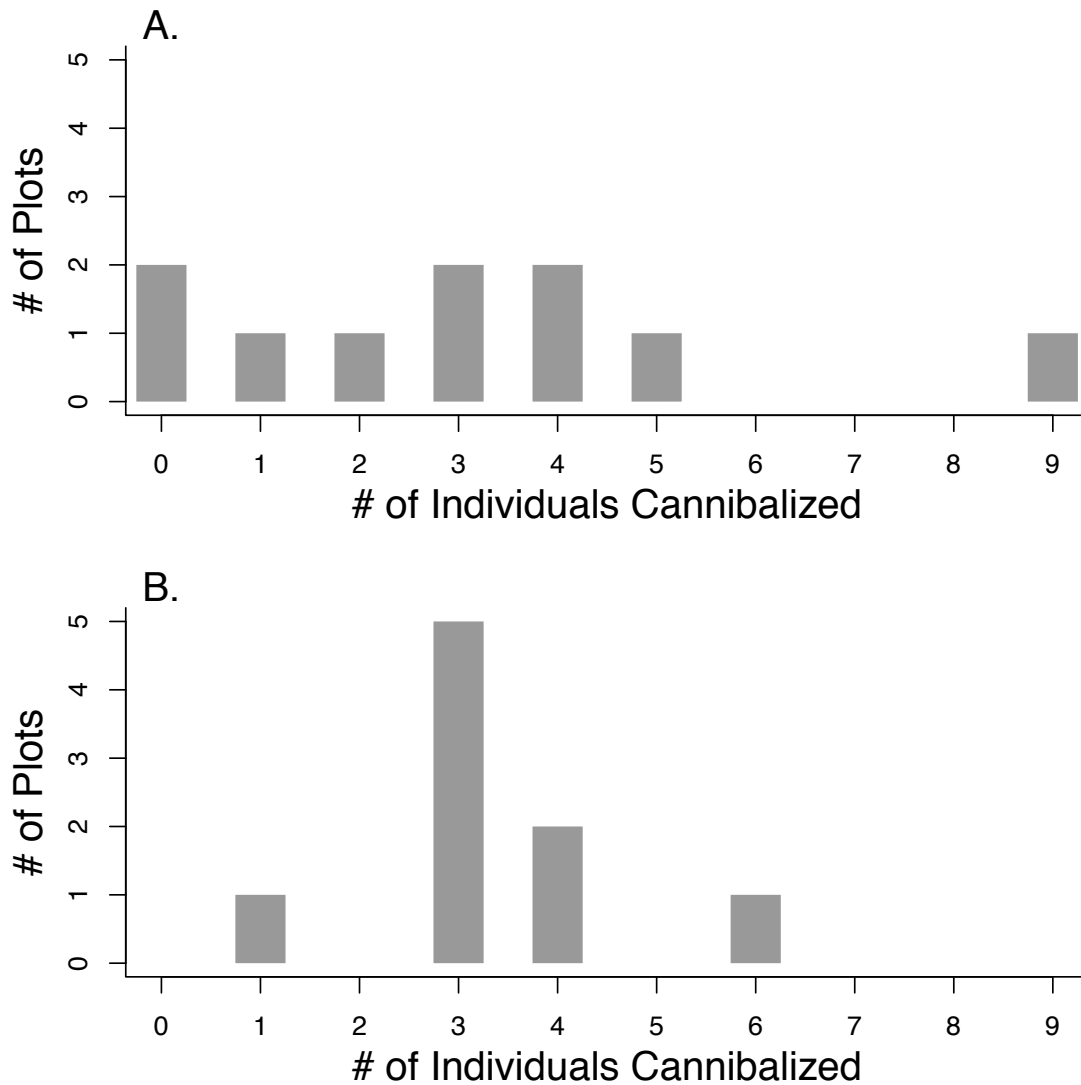


Figure S1: The number of plots along with the number of fourth-instar larvae cannibalized at the end of the field experiment in A) control plots and B) JA-induced plots. The effects of plant induction on number of individuals cannibalized were positive but the 95% Credible Intervals overlapped zero (median [95% CI], 0.07 [-0.428, 0.587]).

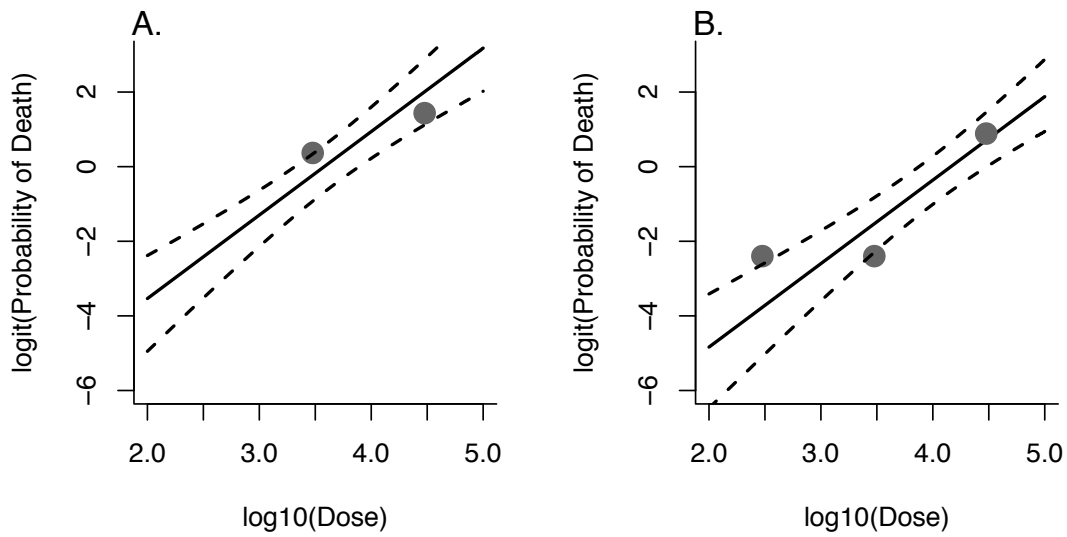


Figure S2: The effects of plant induction on individual-level dose-response to baculovirus infection using the logit transformation of the best-ranked model to fit the data (Table 1). The logit function being $\log\left(\frac{p}{1-p}\right)$, where p is the probability of death from contracting the virus. The figure shows the effects of increasing baculovirus dose given the feeding of baculovirus on A) control leaf tissue and B) jasmonic acid treated leaf tissue. Note that for the control plot that no data are plotted for the $\log_{10}(\text{Dose})$ associated with the 300 occlusion bodies (OBs) treatment. This is because the dose resulted in no infections and the logit of zero is undefined.

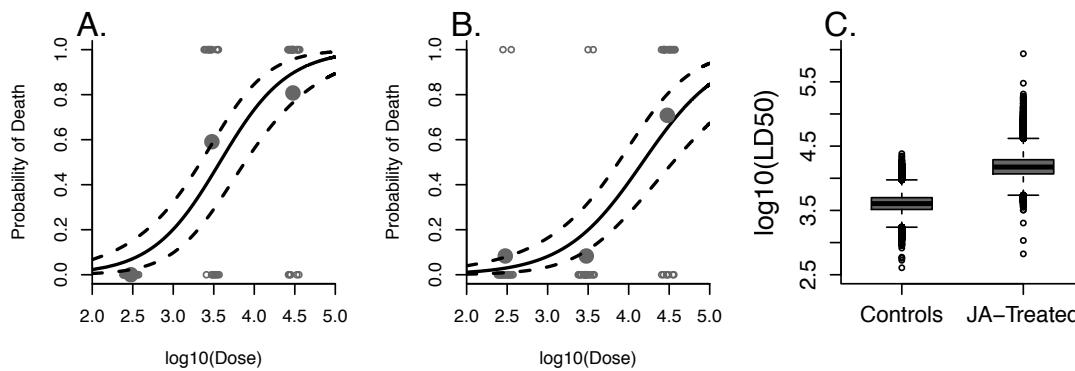


Figure S3: The effects of plant induction on individual-level dose-response to the baculovirus using the second ranked model to fit the data (Table 1). The effects of increasing baculovirus dose on probability of death given the feeding of baculovirus on A) control leaf tissue and B) jasmonic acid treated leaf tissue along with the corresponding C) box plot of the lethal dose at which 50% of the individuals would be expected to die or LD50. For A) and B), the large filled points represent the mean response for each dose and the small open points are the individual data. These data are jittered for ease of presentation.